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On the role of Cro in λ prophage induction

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The lysogenic state of bacteriophage λ is exceptionally stable yet the prophage is readily induced in response to DNA damage. This delicate epigenetic switch is believed to be regulated by two proteins; the lysogenic maintenance promoting protein CI and the early lytic protein Cro. First, we confirm, in the native configuration, the previous observation that the DNA loop mediated by oligomerization of CI bound to two distinct operator regions (O_L and O_R), increases repression of the early lytic promoters and is important for stable maintenance of lysogeny. Second, we show that the presence of the *cro* gene might be unimportant for the lysogenic to lytic switch during induction of the λ prophage. We revisit the idea that Cro's primary role in induction is instead to mediate weak repression of the early lytic promoters.

CI protein | genetic switch | transcription

The λ prophage of *Escherichia coli* can escape lysogeny and enter lytic development by prophage induction (1). Induction is triggered by the host SOS response, which in turn is activated by damage to the host cell DNA. Thus, induction provides a way for the prophage to escape from a challenged or dying host. During lysogeny, the λ lytic genes are repressed by a phage-encoded repressor, the product of the *cI* gene (2). The lytic genes of λ are arranged in a sequential and temporal manner where the expression of one group of genes is required for expression of the next and henceforth. The CI repressor silences all of the lytic genes by preventing transcription from the two earliest lytic promoters. The regulatory region of promoter right (P_R) and promoter left (P_L) each contain an operator, operator right (O_R) and operator left (O_L), respectively, each consisting of three binding sites for CI (reviewed in ref. 3; see also Fig. 1B). In the lysogen, two CI dimers are bound cooperatively to O_R1/O_R2 and O_L1/O_L2 to prevent transcription from P_R and P_L . The CI dimer bound to O_R2 activates transcription from maintenance promoter (P_{RM}) by a direct protein–protein interaction with RNA polymerase (4–7). CI has a lower binding affinity for O_R3 relative to O_L3 and the other operators. Recent studies have shown that O_R3 and O_L3 are simultaneously occupied by CI in lysogens $\approx 60\%$ of the time (8). CI binding to O_R3 excludes RNA polymerase from initiating transcription from P_{RM} , thus *cI* transcription is both positively and negatively autoregulated.

The repression of P_{RM} is enhanced by an O_L –CI– O_R complex, formed by octamerization of the CI dimers bound to O_L1/O_L2 and O_R1/O_R2 (refs. 9–11; see Fig. 1B); thus, the left operator region participates in the regulation of the right operator region and vice versa. The early lytic protein Cro, transcribed from P_R , antagonizes CI after prophage induction (12, 13). Cro binds to the same three sites at O_R and O_L as CI binds, but does so with the opposite affinities of CI. When Cro binds to O_R3 , it prevents transcription of *cI* from P_{RM} , and only at higher concentrations does Cro bind O_R2/O_R1 and repress transcription of P_R . It has been proposed that Cro is also important for the regulation of the switch from lysogenic to lytic growth during λ prophage induction (12–14).

To induce the prophage, P_R and P_L must be derepressed to initiate transcription of the lytic genes. After initiation of the SOS response, this is accomplished through activation of the host RecA coprotease, which binds to CI and promotes autocleavage of CI (15). In a *recA*[–] host, SOS-mediated prophage induction

is defective; the residual induction, caused by fluctuation of CI levels, occurs very rarely, less than once per million cells (16). This frequency is lower than that of mutational inactivation of the *cI* gene (17). Thus, the regulatory network maintaining lysogeny is extremely stable, more stable than the genes encoding it. The high stability is not obtained by precise control of the concentration of CI repressor because single-cell studies have shown that the concentration of repressor in stable lysogens vary greatly from cell to cell (11).

The switch between lysogenic and lytic growth of phage λ was the first genetic switch to be deciphered and the system has contributed immensely to our present understanding of developmental pathways (3). The recent observation that a DNA loop forms between O_L and O_R prompted us to investigate the role of the loop in the switching process of induction. We show here that the interaction between repressors bound at O_L and O_R increases the tolerance of the switch to fluctuations in CI concentration and argues against the role of Cro in the switch that leads to prophage induction.

Materials and Methods

Strains. All strains are derivatives of *E. coli* K12 and are listed in Table 4, which is published as supporting information on the PNAS web site. The parent strain of all λ fusions, NC398, is a bidirectional reporter that carries the luciferase (*luc*) reporter gene on one direction and the β -galactosidase (*lacZ*) reporter gene on the other in the chromosome at the *lac* locus. Any promoter region of interest can be inserted into NC398. All constructs were made by using recombinering (18).

β -Galactosidase Assays. To determine the kinetics of P_R activity upon temperature induction of *cI857* lysogens, cultures were grown in LB (19) at 30°C overnight, diluted 1:200 the next morning, and induced by rapid transfer of the culture flasks to a 42°C water bath when OD₆₀₀ reached 0.3–0.4. At each time point, 1 ml of culture was rapidly transferred to an Eppendorf tube on wet ice. After all time points had been collected and cooled, OD₆₀₀ was recorded and a 0.5-ml sample was immediately mixed with 0.5 ml of Z-buffer (20) containing 25 μ l of chloroform and 25 μ l of 0.1% SDS. β -Galactosidase activity in the sample was then measured as described by Miller (20).

To determine the relationship between growth temperature and activity of P_R in the *cI857* lysogens, cultures were grown at 22°C overnight, diluted 1:200 the following morning, and incubated at the indicated temperatures to reach OD₆₀₀ = 0.3–0.4. Samples were assayed for β -galactosidase activities.

Luciferase Assays. Luciferase activity was measured by using the reagents of the Promega Luciferase Assay System (catalog no. E1500) according to the company's directions (21). Cells were grown and treated as described for the β -galactosidase assays up

Abbreviations: O_R , operator right; O_L , operator left; P_R , promoter right; P_L , promoter left; P_{RM} , maintenance promoter.

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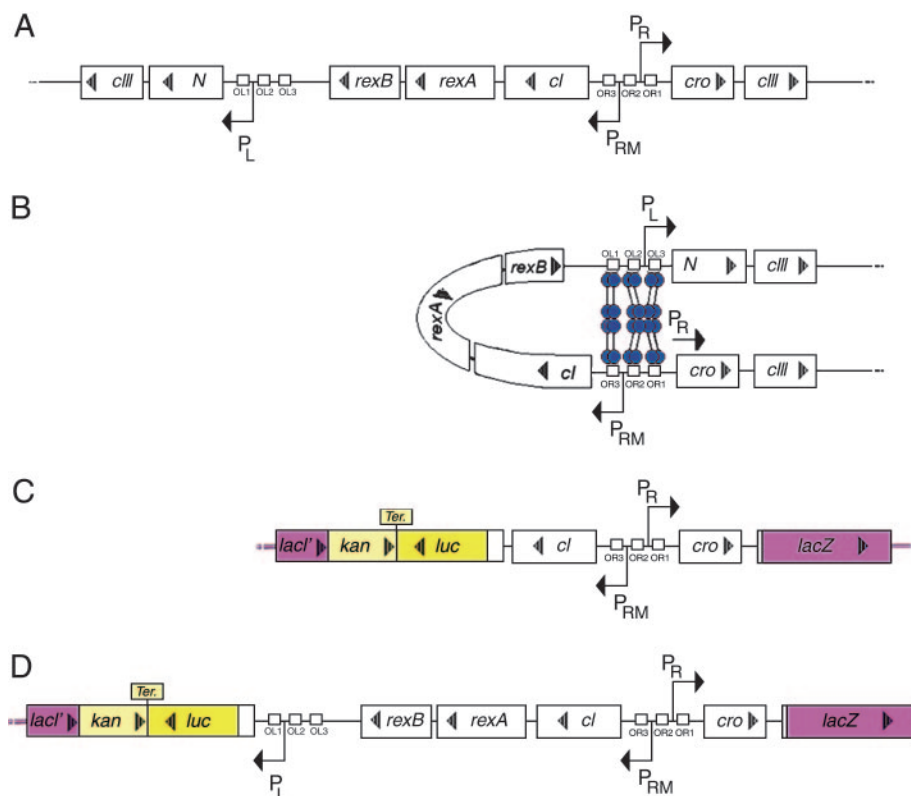


Fig. 1. Genetic map of the immunity region of phage λ showing DNA looping and the various reporter gene fusions. (A) The λ immunity region. (B) The CI dimers bound to O_{L1}/O_{L2} and O_{R1}/O_{R2} can octamerize to create a DNA loop between the operators. (C) To create our reporter strains we placed part of the λ immunity region between the *lac* and *lacZ* reporter genes at the *lac* locus. NC414 and NC415 carry a *rexA::luc* and a *cII::lacZ* fusion. (D) NC416 and NC417 carry an *N::lac* and a *cII::lacZ* fusion.

to the point of addition of lysis buffer. Culture aliquots (0.1 ml) were centrifuged for 2 min at $21,000 \times g$, and pellets were resuspended in 0.4 ml of Cell Culture Lysis Reagent with 2.5 mg/ml BSA and 1.25 mg/ml lysozyme. Twenty microliters of cell lysate was mixed with 0.1 ml of luciferase substrate (Promega Luciferase Assay Reagent, catalog no. E151A), incubated for 120 s and read in EG&G Berthold Lumat LB 9507 single sample luminometer for 10 s. The relative light unit (RLU) was normalized to A_{600} .

Results

The formation of an O_L -CI- O_R regulatory complex containing a DNA loop has been demonstrated to increase repression of P_{RM} and P_R (8, 10, 11). This additional level of regulation is required for the prophage's compensatory response to low doses of DNA damage, which sets the threshold level of DNA damage to which the prophage responds by induction (11). Repressor bound to O_L can also increase repression of the P_R promoter when artificially placed 3,600 bp downstream of the P_R transcription start site, suggesting that the formation of an O_L -CI- O_R complex might also affect transcription from the early lytic promoters (9). To study the effects of the O_L -CI- O_R DNA loop on regulation of the early lytic promoters, we constructed $P_R::lacZ$ fusions by inserting part of the immunity region of λ between two reporter genes at the *lac* locus of *E. coli* as described in *Supporting Text* and Table 4, which are published as supporting information on the PNAS web site, and shown in Fig. 1 C and D. The constructs fuse the *cII* gene with *lacZ*, rendering the expression of β -galactosidase under P_R control. P_R is one of the two early lytic promoters that are repressed in the lysogen but

activated upon induction; thus, production of β -galactosidase in the fusion strains reflects induction of the lytic pathway.

Strains NC416 and NC417 contain the part of the λ immunity region ranging from the AUG initiation codon of gene *N* gene on the left to the 30th codon of *cII* on the right. These strains retain both the left and the right operator regions (Fig. 1D). In strains NC414 and NC415, only the right operator region is retained; the strains carry the λ immunity region from the AUG initiation codon of the *rexA* gene on the left to the 30th codon of *cII* on the right (Fig. 1C). The AUG codon of *N* or *rexA* is fused to the 2nd codon of the *lac* reporter gene, while the 30th codon of *cII* is fused to the 7th codon of *lacZ*. In strains NC414 and NC415, luciferase is expressed from P_{RM} , whereas in NC416 and NC417, the reporter is under the control of the lytic P_L promoter. The fusions also vary with respect to the *cro* gene. NC414 and NC416 carry the wild-type *cro* gene, whereas NC415 and NC417 carry a nonfunctional missense *cro27* allele (12). Together, the four strains represent the four possible combinations of the O_L^+ or ΔOL and *cro*⁺ or *cro*⁻ genotypes.

Table 1. Activity of β -galactosidase in repressed lysogens

Strain (relevant genotype)	β -Galactosidase activity, Miller units
NC414 (ΔO_L , <i>cro</i> ⁺)	21 (3.1)
NC415 (ΔO_L , <i>cro</i> ⁻)	15 (1.1)
NC416 (O_L^+ , <i>cro</i> ⁺)	4 (1.2)
NC417 (O_L^+ , <i>cro</i> ⁻)	4 (1.3)

The O_L -CI- O_R complex increases repression of P_R 4-fold. Samples were grown at 30°C and assayed for β -galactosidase activity in early log phase as described in *Materials and Methods*. The averages of four independent experiments are shown. Standard deviations are given in parentheses.



Fig. 2. Morphology of NC414–417 after growth at 30°C on MacConkey lactose agar (20). The colonies were grown for 42 h after restreaking from single colonies.

The O_L - CI - O_R Complex Increases Repression of P_R in the Native Prophage Configuration. The expression of β -galactosidase in log phase NC414, NC415, NC416, and NC417 cultures growing at 30°C under repressed conditions is shown in Table 1. It is evident that the presence of the O_L region increases repression of P_R when the immunity region is located in the chromosome in its native configuration confirming the results obtained in multicopy systems (8–10). Note that our O_L -deletion also removes the *rex* genes between O_L and *cI*. We addressed the possibility that the decreased repression in the O_L -deletion is not because of the absence of *rex* and demonstrated that the presence or absence of *rex* genes in the O_L^+ background does not measurably affect repression of P_R at temperatures ranging from 30°C to 37°C (data not shown). In the absence of functional Cro, the β -galactosidase activity is ≈ 4 -fold higher in the absence of the left operator region than in its presence, reflecting that O_L increases the repression of P_R 4-fold. The 4-fold increase in P_R activity with O_L deleted is essentially the same as when CI is supplied at a constant level in trans (8–10), showing that O_L is increasing the efficiency of P_R repression at the normal lysogenic concentration of CI by >4 -fold. We noted that in the absence of the left operator region, P_R activity is decreased $\approx 30\%$ in the *cro*[−] mutant (NC415) as compared to *cro*⁺ (NC414). This observation could either reflect a uniformly elevated expression from P_R in the *cro*⁺ cells in comparison with the *cro*[−] cells, or it could reflect that a fraction of the *cro*⁺ population has switched to

the “lytic” state where P_R is not repressed by CI . The latter possibility is supported by the observation that, when the four strains are grown on MacConkey lactose agar plates, red colonies appear more frequently among NC414 (O_L^- *cro*⁺) cells than among NC415, NC416, and NC417 cells (Fig. 2). The red color indicates that these cells are producing higher levels of β -galactosidase, which metabolizes lactose. However, when both O_L and O_R regions are present, we were unable to detect any difference between *cro*⁺ and *cro*[−] in P_R expression (Table 1, compare NC416 and NC417). This observation could indicate that the interaction between O_L and O_R prevents Cro from switching the prophage from the lysogenic to the lytic state. To examine this hypothesis further, we studied the activity of P_R at low repressor concentrations. The *cI* allele present in our reporter strains is the *cI857* allele, which encodes a temperature-sensitive repressor protein (22). The *cI857* allele behaves as wild-type *cI* at 30°C, whereas there is essentially no repressor activity at temperatures above 40°C. Thus, a range of repressor activities can be obtained by growing the strains at a range of temperatures between the permissive and restrictive temperatures.

Role of Cro in the Switch Process? The expression of *lacZ* as a function of growth temperature for NC414, NC415, NC416, and NC417 is shown in Fig. 3. If *cro* is critical for the switch (14), we expect that derepression of P_R will occur at lower temperatures in the *cro*⁺ than in the *cro*[−] cells, suggesting that Cro can sway the switch from lysogenic to lytic growth at intermediate concentrations of CI . We obtained that result for the strains that do not carry the left operator region: NC414(*cro*⁺) and NC415(*cro*[−]) (Fig. 3). However, the expected pattern was not seen in the strains that carry both the left and the right operator regions. Although the role of *rex* genes, if any, in the switch process remains to be investigated in strains containing both operator regions, no difference in β -galactosidase levels were observed between NC416(*cro*⁺) and NC417(*cro*[−]) cells. In fact, the derepression of P_R as a function of growth temperature appears identical for NC415, NC416, and NC417, suggesting that the DNA loop between the left and right operator region renders the switch insensitive to the presence of the *cro*⁺ gene.

The Effect of Cro Mediated Repression of P_R Transcription in the Presence and Absence of O_L . We investigated whether the left operator region had any influence on the regulation of P_R in the absence of functional CI . We inactivated CI by shifting growing cultures from 30°C to 42°C and followed the kinetics of P_R derepression by monitoring β -galactosidase activity at various times

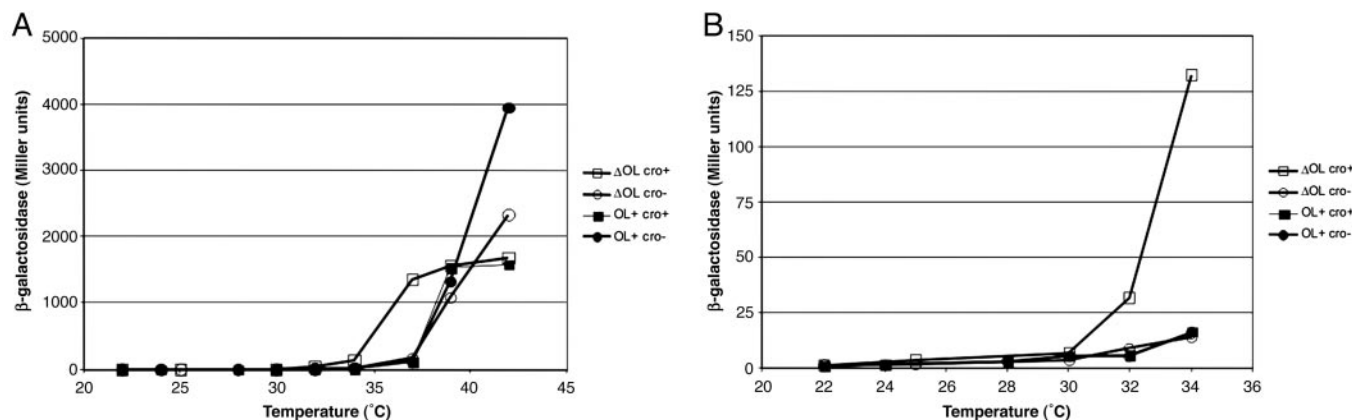


Fig. 3. β -Galactosidase activity as a function of growth temperature. (A) The activity of β -galactosidase at growth temperatures in the range between 22°C and 42°C. (B) A magnification of the left part of A, showing the activity of β -galactosidase at low temperatures. Open squares, NC414 (ΔO_L *cro*⁺); open circles, NC415 (ΔO_L *cro*[−]); filled squares, NC416 (O_L^+ *cro*⁺); filled circles, NC417 (O_L^+ *cro*[−]). Assays were performed after 18 h of growth at the indicated temperatures. The absolute values reported here are different from those presented in Fig. 4 and Table 2, in which cases log phase cells were used. The average activity of two to four independent cultures are shown in Miller units.

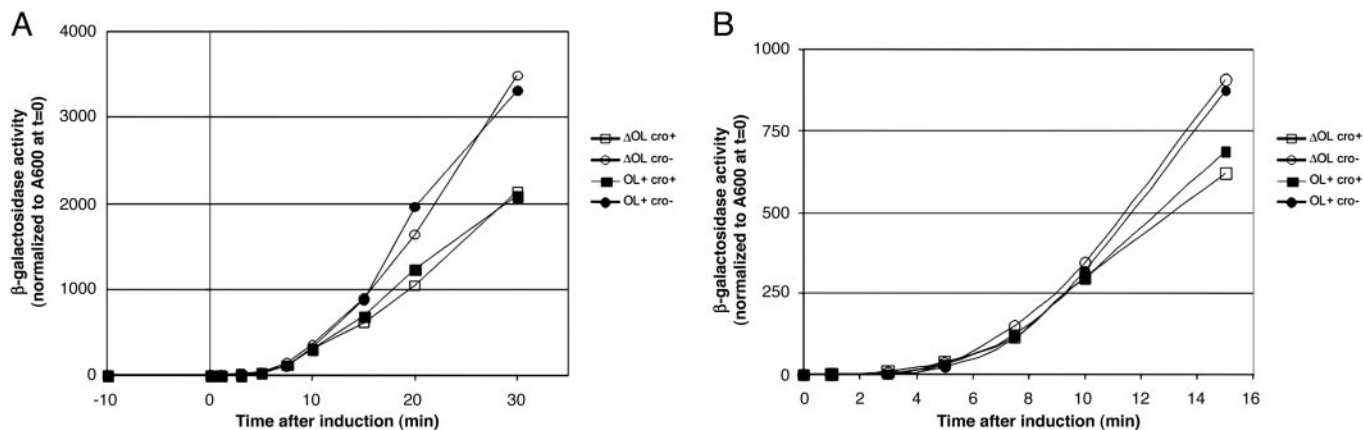


Fig. 4. β -Galactosidase activity after thermal induction. (A) The β -galactosidase activities of NC414, NC415, NC416, and NC417 at various time points after shift of growth temperature from 30°C to 42°C normalized to A_{600} at the time of the temperature shift. Open squares, NC414 (ΔO_L^- cro^+); open circles, NC415 (ΔO_L^- cro^-); filled squares, NC416 (O_L^+ cro^+); filled circles, NC417 (O_L^+ cro^-). The β -galactosidase activities of cultures maintained at 30°C has been subtracted from all values. (B) A magnification of the left part A. The average activity of two to four independent cultures is shown.

after the temperature switch. The activities of β -galactosidase as a function of time after induction for NC414, NC415, NC416, and NC417 are shown in Fig. 4. For NC414 and NC416, which carry cro^+ , it can be seen that ≈ 10 min after induction, Cro presumably has accumulated to a level where it binds O_R1 and/or O_R2 and represses P_R , causing the cro^+ curves to diverge from the cro^- curves (NC415 and NC417). It took equally long for Cro to accumulate to a concentration where it negatively autoregulates P_R in NC414 (ΔO_L) and NC416 (O_L^+), and the degree of Cro-mediated repression of P_R was similar in the two strains (Table 2). Therefore, we saw no indication that the left operator region participated in regulation of P_R by Cro when CI was absent as expected (8), consistent with the hypothesis that O_L participates in regulation at O_R through the CI-mediated DNA loop (10).

Repression by Cro. It was suggested earlier on that the primary role of Cro in prophage induction is to repress the early lytic promoters, P_R and P_L , as well as the maintenance promoter P_{RM} . The relationship between CI and Cro is that CI is a strong repressor specialized for complete turnoff of lytic functions, whereas Cro is a weak repressor functioning in a partial turn down of the early lytic promoters to allow progression into the late lytic phase (23, 24). Our unique divergent reporter constructs allowed us to measure the degree of repression of P_R , P_L , and P_{RM} by CI and Cro in this study. To measure the degree of repression of P_L by Cro, we compared the luciferase activity of NC416 ($N::luc$, cro^+) and NC417 ($N::luc$, cro^-) at 42°C, where CI857 is inactive (Table 3). Likewise, to measure the degree of repression of P_{RM} by Cro, we compared luciferase in NC414 ($rexA::luc$, cro^+) and NC415 ($rexA::luc$, cro^-) at 42°C (Table 3). The degree of repression of P_L and P_R by CI in the presence of both the left and right operator regions was estimated by comparing the activity of the respective reporter genes in NC417 ($P_R::lacZ$, $P_L::luc$, cro^-) grown at 30°C, where CI857 is active, with NC417

grown at 42°C, where CI857 is inactive (Table 3). The luciferase protein is inherently heat sensitive and forms inactive aggregates after temperature shifts, but enzyme activity is restored with the aid of host chaperones after a few minutes at the high temperature (25). We observed full restoration of luciferase activity ≈ 15 min after shifting from 30°C to 42°C in a reporter containing wild-type CI (not heat-sensitive; strain WA5) (data not shown). Luciferase activities reported here were measured 60 min after the temperature shift. We observed that, in the absence of CI, Cro represses the P_L promoter ≈ 1.7 -fold (Table 3), although a stronger repression was reported when a fusion in which transcription terminators were present between P_L and a reporter gene ($galK$) was used (26). In the current bidirectional reporter setup, Cro repressed P_{RM} promoter ≈ 2.5 -fold. (Table 3).

Discussion

Repression of P_R by the O_L -CI- O_R Complex in its Native Configuration.

Tight repression of P_R is believed to be vital for the maintenance of lysogeny, because otherwise Cro would be produced, bind to O_R3 , and repress P_{RM} , turning off cI transcription (27). Mathematical modeling studies of the lysis-lysogeny switch have concluded that CI bound at O_R cannot repress Cro production sufficiently to stably maintain the lysogen unless there are additional levels of repression of P_R (16, 28). We have confirmed that an interaction between the left and right operator region increases CI-mediated repression of P_R 4-fold. Thus, our data are consistent with that of Revet *et al.* (9) and Dodd *et al.* (8) in suggesting that the DNA loop between O_L and O_R constitutes the missing level of regulation. Our data also support the model of Dodd *et al.* (8) to explain that the means by which the presence of O_L allows tighter repression of P_R at physiological CI concentrations. The extra level of cooperativity added by the formation of a CI octamer increases the binding affinity of CI to the involved operator sites, thus lowering the critical concentration of CI needed to maintain lysogeny.

O_L Contributes to Repression of Cro Synthesis. According to the classic model, Cro's major role during induction is to bind O_R3 and thereby inhibit cI expression from P_{RM} (14, 27). If a Cro-mediated reduction in cI expression were important for effective induction of the early lytic promoters, we would expect a $cI857$ cro^- prophage grown at intermediate temperatures to contain more repressor than the corresponding cro^+ prophage, and thus require inactivation of a larger fraction of the repressor proteins to effectively derepress P_R . Instead, we observe the same amount of derepression of P_R at the same growth temperature in cro^+ (NC416) and cro^- (NC417),

Table 2. Autorepression by Cro after prophage induction

Strain (relevant genotype)	Slope, * Miller units/min	cro^-/cro^+
NC415 (ΔO_L cro^-)	120	1.62
NC414 (ΔO_L cro^+)	74	
NC417 (O_L^+ cro^-)	147	1.65
NC416 (O_L^+ cro^+)	89	

β -Galactosidase activities were assayed as described in *Materials and Methods*.

*Slope of the linear part of the induction curves shown are from Fig. 4.

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